

Genomic DNA Preamplification Using the Advanta CFTR NGS Preamp Reagent Kit

IMPORTANT Before using this quick reference, read and understand the detailed instructions and safety guidelines in the Advanta™ CFTR NGS Library Preparation on the LP 48.48 IFC with Juno™ Protocol (PN 101-6270) or the Advanta CFTR Targeted Library Preparation on the Juno LP 192.24 IFC Protocol (PN 101-6212).

Prepare the Preamplification Reactions

IMPORTANT Preamplified and non-preamplified samples can be run on the same IFC. However, libraries for each must be generated separately and normalized before combining and sequencing. Failure to independently generate and normalize the libraries will result in unequal sequencing read distributions between preamplified and non-preamplified samples.

- 1 Manually flick the bottom of the TSP DNA Polymerase tube with your forefinger to mix the enzyme (**do not vortex**). Vortex all other reagents for 20 sec, and then briefly centrifuge all reagents before use.

- 2 In a DNA-free hood, prepare the preamplification pre-mix in a 1.5 mL tube as shown in Table 1.

Table 1. Preamplification pre-mix. Combine and mix in order shown.

Component	Volume per Reaction (µL)	Volume for 96 Reactions (µL)*
1 PCR Water (PN 100-5941)	1.8	216
2 4X TSP Master Mix (PN 101-3055)	1.25	150
3 TSP Sample Loading Reagent v2 (PN 101-7634)	0.25	30
4 TSP DNA Polymerase (PN 101-0995)	0.2	24
5 Advanta CFTR NGS Preamp Pool (PN 101-7279)	0.5	60
Total	4.0	480

*Includes overage.

- 3 Mix the preamplification pre-mix by briefly vortexing, and then centrifuge to bring down the contents.
- 4 Prepare the preamplification reaction mixes per sample as shown in Table 2 and Figure 1.
 - a Using an 8-channel pipette, transfer preamplification pre-mix into the wells of a 96-well PCR plate.
 - b Remove the plate from the DNA-free hood and add DNA samples to each well containing pre-mix, as shown in Figure 1.

Table 2. Preamplification reaction mix

Component	Volume per Reaction (µL)
Preamplification pre-mix (see Table 1)	4.0
Genomic DNA (gDNA), 5–30 ng/µL	1.0
Total	5.0

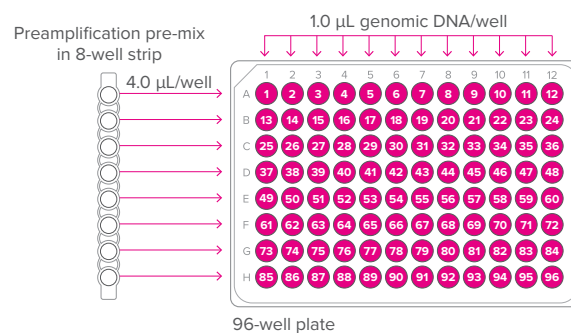


Figure 1. Preparation of preamplification reaction mix

- 5 Seal the plate using an adhesive seal.
- 6 Mix the reactions by briefly vortexing, and then centrifuge at 3,000 x g for 1 min to bring down contents.
- 7 Place the plate in a standard thermal cycler and cycle using the following thermal protocol:

Temperature	Time	Cycles	Description
95 °C	15 min		Hold
95 °C	15 sec	14	Denaturation
60 °C	8 min		Annealing/extension
4 °C	∞		Hold

Cycling time is approximately 2 hr and 15 min.

- 8 After cycling, dilute the product as shown in Table 3.

Table 3. Preamplification product dilution

Component	Volume per Reaction (µL)
Preamplification product	2
DNA Dilution Reagent (PN 100-9167)	38
Total	40

- 9 Seal the plate using an adhesive seal.
- 10 Mix the diluted products by briefly vortexing, and then centrifuge at 3,000 x g for 1 min to bring down contents.
- 11 Use 2.5 µL of the diluted preamplified gDNA as a template for preparing the sample mixes.

STOPPING POINT The diluted preamplification products can be stored at 2–8 °C if used within 1 week, or stored at –20 °C for later use.

For technical support visit fluidigm.com/support.

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